* **Sectioning**
  1. Pull block from -80oC freezer.
  2. Place in cryostat, and allow to equilibrate for 30 minutes.
  3. Set cryostat to cut slices of 7µm thickness.
* **Pre-Staining**
  1. Use a hydrophobic pen to draw a 0.75" x 0.75" square around each slice on the slides. Make multiple passes around the square to ensure the tissue is enclosed adequately.
* **Staining**
  1. Hematoxylin: **\*2 experimental conditions\***
     + **5-minute** Hematoxylin exposure
       - Apply 170 μL of Hematoxylin on each slice, and allow to sit for 5 minutes.
     + **1-minute** Hematoxylin exposure
       - Apply 170 μL of Hematoxylin on each slice, and allow to sit for 1 minute.
  2. Rinse slides in tap water.
  3. Apply 170 μL of Bluing Reagent on each slice, and allow to sit for 30 seconds.
  4. Rinse slides in DI water.
  5. Dip slides in 95% ethanol for 5 seconds.
  6. Apply 170 μL of Congo Red on each slice, and allow to sit for 1 minute.
  7. Dip slides once in 2 troughs of 100% ethanol (2 dips total).
  8. Place slides in a trough containing xylenes, and allow to sit for 1 minute.
  9. Allow slides to dry for ~5 minutes prior to proceeding.
* **Post-Staining**
  1. Gently sway slides, one at a time, in a trough containing 100% ethanol for 20 seconds, followed by the same movement in a trough containing xylenes for 20 seconds.
  2. Coverslip with paramount.
  3. Allow to dry in the fume hood for ~30 minutes prior to further examination.